



Clinical randomized comparison of Medetomidine and Xylazine for isoflurane balanced anesthesia in horses

Wiederkehr, Alexandra ; Barbarossa, Andrea ; Ringer, Simone K ; Jörger, Fabiola Binia ; Bryner, Marco ; Bettschart-Wolfensberger, Regula

Abstract: Introduction: To assess drug plasma levels, preanesthetic sedation, cardiopulmonary effects during anesthesia and recovery in horses anesthetized with isoflurane combined with medetomidine or xylazine. Study design: Prospective blinded randomized clinical study. Animals: Sixty horses undergoing elective surgery. Methods: Thirty minutes after administration of antibiotics, flunixin meglumine or phenylbutazone and acepromazine horses received medetomidine $7 \mu\text{g kg}^{-1}$ (group MED) or xylazine 1.1 mg kg^{-1} (group XYL) slowly intravenously (IV) and sedation was assessed 3 min later. Anesthesia was induced with ketamine/diazepam and maintained with isoflurane in oxygen/air and medetomidine $3.5 \mu\text{g kg}^{-1} \text{ h}^{-1}$ or xylazine $0.69 \text{ mg kg}^{-1} \text{ h}^{-1}$. Ringer's acetate $10 \text{ mL kg}^{-1} \text{ h}^{-1}$ and dobutamine were administered to maintain normotension. All horses were mechanically ventilated to maintain end-tidal carbon dioxide pressures at $45 \pm 5 \text{ mmHg}$ ($5.3\text{--}6.7 \text{ kPa}$). Heart rate (HR), invasive arterial blood pressures, inspired and expired gas compositions, pH, arterial blood gases, electrolytes, lactate and glucose were measured. For recovery all horses received intramuscular morphine 0.1 mg kg^{-1} and medetomidine $2 \mu\text{g kg}^{-1}$ or xylazine 0.3 mg kg^{-1} IV. Recovery was timed and scored using three different scoring systems. Plasma samples to measure medetomidine and xylazine concentrations were collected at predetermined timepoints. Repeatedly measured parameters were analyzed using a two-way repeated-measures analysis of variance for differences between groups and over time; $p < 0.05$ was considered statistically significant. Results: Mean arterial blood pressures (MAP) stayed within normal ranges but were higher ($p = 0.011$) in group XYL despite significant lower dobutamine doses ($p = 0.0003$). Other measured parameters were within clinically acceptable ranges. Plasma levels were at steady state during anesthesia (MED 2.194 ± 0.073 ; XYL $708 \pm 18.791 \text{ ng mL}^{-1}$). During recovery lateral recumbency (MED 42.7 ± 2.51 ; XYL $34.3 \pm 2.63 \text{ min}$; $p = 0.027$) and time to standing (MED 62.0 ± 2.86 ; XYL $48.8 \pm 3.01 \text{ min}$; $p = 0.002$) were significantly shorter in group XYL compared to group MED. Recovery scores did not differ significantly between groups. Conclusion and Clinical Relevance: In horses anesthetized with isoflurane and medetomidine or xylazine, xylazine maintained higher MAP, reduced the dobutamine consumption and recovery time, whilst overall recovery quality was unaffected.

DOI: <https://doi.org/10.3389/fvets.2021.603695>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-202822>

Journal Article

Published Version



The following work is licensed under a Creative Commons: Attribution 4.0 International (CC BY 4.0) License.

Originally published at:

Wiederkehr, Alexandra; Barbarossa, Andrea; Ringer, Simone K; Jörger, Fabiola Binia; Bryner, Marco; Bettschart-Wolfensberger, Regula (2021). Clinical randomized comparison of Medetomidine and Xylazine for isoflurane balanced anesthesia in horses. *Frontiers in Veterinary Science*, 8:603695.

DOI: <https://doi.org/10.3389/fvets.2021.603695>



Clinical Randomized Comparison of Medetomidine and Xylazine for Isoflurane Balanced Anesthesia in Horses

Alexandra Wiederkehr^{1*}, Andrea Barbarossa², Simone K. Ringer¹, Fabiola B. Jörgen¹, Marco Bryner³ and Regula Bettschart-Wolfensberger¹

¹ Section Anesthesiology, Department of Clinical Diagnostics and Services, Vetsuisse Faculty, University of Zurich, Zurich, Switzerland, ² Department of Veterinary Medical Sciences, University of Bologna, Bologna, Italy, ³ Clinic for Equine Surgery, Equine Department, Vetsuisse Faculty, University of Zurich, Zurich, Switzerland

OPEN ACCESS

Edited by:

Klaus Hopster,
University of Pennsylvania,
United States

Reviewed by:

Angela Briganti,
University of Pisa, Italy
Dario Floriano,
University of Pennsylvania,
United States

*Correspondence:

Alexandra Wiederkehr
alexandra.wiederkehr@uzh.ch

Specialty section:

This article was submitted to
Veterinary Surgery and
Anesthesiology,
a section of the journal
Frontiers in Veterinary Science

Received: 07 September 2020

Accepted: 22 March 2021

Published: 20 April 2021

Citation:

Wiederkehr A, Barbarossa A, Ringer SK, Jörgen FB, Bryner M and Bettschart-Wolfensberger R (2021) Clinical Randomized Comparison of Medetomidine and Xylazine for Isoflurane Balanced Anesthesia in Horses. *Front. Vet. Sci.* 8:603695. doi: 10.3389/fvets.2021.603695

Introduction: To assess drug plasma levels, preanesthetic sedation, cardiopulmonary effects during anesthesia and recovery in horses anesthetized with isoflurane combined with medetomidine or xylazine.

Study design: Prospective blinded randomized clinical study.

Animals: Sixty horses undergoing elective surgery.

Methods: Thirty minutes after administration of antibiotics, flunixin meglumine or phenylbutazone and acepromazine horses received medetomidine 7 $\mu\text{g kg}^{-1}$ (group MED) or xylazine 1.1 mg kg^{-1} (group XYL) slowly intravenously (IV) and sedation was assessed 3 min later. Anesthesia was induced with ketamine/diazepam and maintained with isoflurane in oxygen/air and medetomidine 3.5 $\mu\text{g kg}^{-1} \text{ h}^{-1}$ or xylazine 0.69 $\text{mg kg}^{-1} \text{ h}^{-1}$. Ringer's acetate 10 $\text{mL kg}^{-1} \text{ h}^{-1}$ and dobutamine were administered to maintain normotension. All horses were mechanically ventilated to maintain end-tidal carbon dioxide pressures at 45 ± 5 mmHg (5.3–6.7 kPa). Heart rate (HR), invasive arterial blood pressures, inspired and expired gas compositions, pH, arterial blood gases, electrolytes, lactate and glucose were measured. For recovery all horses received intramuscular morphine 0.1 mg kg^{-1} and medetomidine 2 $\mu\text{g kg}^{-1}$ or xylazine 0.3 mg kg^{-1} IV. Recovery was timed and scored using three different scoring systems. Plasma samples to measure medetomidine and xylazine concentrations were collected at predetermined timepoints. Repeatedly measured parameters were analyzed using a two-way repeated-measures analysis of variance for differences between groups and over time; $p < 0.05$ was considered statistically significant.

Results: Mean arterial blood pressures (MAP) stayed within normal ranges but were higher ($p = 0.011$) in group XYL despite significant lower dobutamine doses ($p = 0.0003$). Other measured parameters were within clinically acceptable ranges. Plasma levels were at steady state during anesthesia (MED 2.194 ± 0.073 ; XYL 708 ± 18.791 ng mL^{-1}). During recovery lateral recumbency (MED 42.7 ± 2.51 ; XYL 34.3 ± 2.63 min; $p = 0.027$) and time to standing (MED 62.0 ± 2.86 ; XYL 48.8 ± 3.01 min; $p = 0.002$)

were significantly shorter in group XYL compared to group MED. Recovery scores did not differ significantly between groups.

Conclusion and Clinical Relevance: In horses anesthetized with isoflurane and medetomidine or xylazine, xylazine maintained higher MAP, reduced the dobutamine consumption and recovery time, whilst overall recovery quality was unaffected.

Keywords: anesthesia, horse, isoflurane, medetomidine, α_2 -adrenergic agonists, equine, equisedative, recovery

INTRODUCTION

Volatile anesthetic agents are commonly used in horses undergoing surgery; however, they cause cardio-respiratory depression (1) and they contribute to the equine anesthesia-associated morbidity and mortality (2, 3). Therefore, a balanced anesthetic protocol with partial intravenous anesthesia (PIVA) using one or more complementary agents is a common concept in modern equine general anesthesia (4–6). The aim of PIVA is to improve analgesia and to reduce the amount of the volatile anesthetic agents (7), in order to maintain good intraoperative cardiopulmonary function followed by smooth and coordinated recovery (4, 5, 8). Various α_2 -adrenergic agonists constant rate infusions (CRI) have been used in anesthetized horses for this purpose (9–13).

Half-lives of medetomidine (4.14 min) (14) and xylazine (5.9 min) (15) are relatively short and onset of action of the drugs is quick. Both drugs have been shown to result in constant plasma levels when administered as CRIs preceded by a bolus (15, 16).

Medetomidine, a potent, short acting, highly selective and specific α_2 -adrenergic agonist [selectivity ratio (α_2 : α_1) 1,620:1] (17), has been shown to reduce the minimum alveolar concentration of isoflurane and provide potent analgesia (5, 18).

The reported side-effects after injection of a single IV bolus (dose) of medetomidine in non-anesthetized horses and ponies are brady-arrhythmias, increase in systemic vascular resistance and arterial blood pressure, and decrease of cardiac output (14, 19, 20). However, these effects are transitory and are not worsened by a following infusion (19). Even though it is not registered for horses in Europe or the United States, there are numerous reports about its use in equine clinical practice (6, 18, 21, 22). The administration of medetomidine CRI produced better recoveries than lidocaine or S-ketamine CRIs in isoflurane anesthetized horses (1, 23).

Xylazine, another α_2 -adrenergic agonist registered for horses also provides sedation, analgesia and muscle relaxation (24). It is the least selective α_2 -agonist [selectivity ratio (α_2 : α_1) 160:1] and widely used as a premedicant (16, 25), but reports about its use as CRI in balanced anesthesia regimes are sparse (11).

Cardiopulmonary effects following xylazine bolus administration are also similar to other α_2 -adrenergic agonists, but during CRI the effects are minimal in particular if compared to romifidine CRI (26). In comparison to medetomidine cardiovascular effects of xylazine have been

demonstrated in ponies to be less and shorter lasting (27). However, it remains to be tested if the dose rates investigated were equipotent. A bolus administration of IV xylazine at 0.5 and 1.0 mg kg⁻¹ reduced isoflurane MAC by 25 and 34%, respectively (28).

The aim of the present study was to compare the intraoperative cardiovascular effects and recovery of xylazine vs. medetomidine isoflurane PIVA in horses undergoing elective surgery. The hypothesis of our study was that 7 μ g kg⁻¹ medetomidine followed by 3.5 μ g kg⁻¹ h⁻¹ results in similar effects as 1.1 mg kg⁻¹ xylazine followed by 0.69 mg kg⁻¹ h⁻¹.

MATERIALS AND METHODS

Study Design and Animals

This prospective randomized blinded clinical study was performed with the ethical approval of the local committee for animal experimentation (ZH176/17). Written owner's consent was obtained.

Sixty client-owned horses of various breeds presented for elective surgeries were included in the study. This number was considered adequate to ascertain that the doses of medetomidine and xylazine tested were suitable regarding their influence on the concentration of isoflurane necessary to maintain anesthesia. In order to detect if isoflurane is increased from 1.1 ± 0.135 to 1.2% with a power of 0.8 and an alpha error of 0.05, 29 horses in each group are necessary. Inclusion criteria were body weight (≥ 200 kg), age (2–20 years), physical status American Society of Anesthesiologists 1 or 2. Exclusion criteria were surgery of the head, need of assisted recovery and general anesthesia in the previous 6 months. Food, but not water, was withheld 8–12 h prior to anesthesia. Each patient was randomly assigned to either group MED (medetomidine) or XYL (xylazine) at the beginning of the experiment, using one of sixty previously prepared opaque envelopes (containing group allocation), opened by a person who was not involved in the study. Demographic data was recorded on a separate data sheet (AW). All anesthetic procedures were performed by the same experienced anesthetist (AW), who was unaware of the treatment at any time until the end of the study.

Study Protocol

Prior to sedation, the skin over one jugular vein was clipped and aseptically prepared. After infiltration with mepivacaine 2% (Mepivacain HCL 2%; Sintetica S.A., Switzerland), a 14 gauge catheter (Secalon-T with FlowSwitch; Argon Critical Care Systems, Singapore) was placed.

At 30–60 min before induction of anesthesia, patients in both groups received 30'000 I.E. kg^{-1} of penicilline Na^+ (Penicillin Natrium Streuli ad us. vet.; Streuli Pharma AG, Switzerland), 6.6 mg kg^{-1} of gentamicine (Genta 10%; CP-Pharma, Switzerland) intravenously (IV). Horses undergoing soft tissue surgeries received flunixin meglumine 1.1 mg kg^{-1} IV (Flunixinim ad us. vet.; Graeb AG, Switzerland) and horses undergoing orthopedic surgeries received phenylbutazone 4 mg kg^{-1} IV (Butadion ad us. vet.; Streuli Pharma AG, Switzerland). All horses received 0.03 mg kg^{-1} of acepromazine (Prequillan ad us. vet.; Arovet AG, Switzerland) IM. Immediately before induction of anesthesia, horses were sedated with medetomidine 7 $\mu\text{g kg}^{-1}$ IV (Medetor[®] ad us. vet.; Virbac AG, Switzerland) or xylazine 1.1 mg kg^{-1} IV (Xylazin 2%; Streuli Pharma AG, Switzerland) depending on group allocation. Medication were prepared by a veterinarian not involved in the study and diluted with sodium chloride to a total volume of 40 mL. One-third of the dose was administered as a bolus in the stable, then the mouth was rinsed and the horse walked to the induction area, where the remaining two-thirds of sedative were injected IV slowly over 2 min. Three minutes after the end of injection, depth of sedation was assessed according to criteria published by Taylor et al. (29): where sedation is indicated by head height lower than withers (eyes as benchmark), atonic lower lip, no reaction to snapping fingers. If one or more than one condition indicating sedation was not present, a supplemental dose of medetomidine 1 $\mu\text{g kg}^{-1}$ or xylazine 0.16 mg kg^{-1} was administered IV, depending on group allocation. Sedation was re-evaluated after 3 min. This procedure was repeated if necessary. The total amount of time required for sedation was recorded.

Anesthesia was induced with IV diazepam 0.02 mg kg^{-1} IV (Valium[®] 5 mg; Roche Pharma AG, Switzerland) and ketamine 2.2 mg kg^{-1} (Ketanarkon 100 ad us. vet.; Streuli Pharma AG, Switzerland) mixed in one syringe. This was considered as anesthesia time point zero (t_0). Following induction of anesthesia and tracheal intubation (silicone tubes, internal diameter 18–30 mm), the horses were hoisted onto a padded surgery table and the endotracheal tube attached to a large animal circle system (Tafonius; Hallowell Engineering & Manufacturing Corp., MA, USA). Isoflurane was delivered using a precision vaporizer (Tec 7, Rothacher Medical GmbH, Heitenried, Switzerland). The carrier gas for isoflurane (IsoFlo[®], Provect AG, Switzerland) delivery consisted of a mixture of oxygen and air with an initial inspiratory fraction of oxygen ($F_{\text{I}}\text{O}_2$) of 0.45–0.55. Every 5 min, the depth of anesthesia was assessed and isoflurane supply readjusted based on judgement by the main investigator, who was unaware of the treatment. Isoflurane was delivered to the end-tidal concentration ($F_{\text{E}}\text{Iso}$) required to keep the patient immobile, to prevent nystagmus and hypertension, to provide muscle relaxation and to allow evocation of sluggish palpebral reflexes. Mechanical ventilation was commenced (tidal volume 10–12 mL kg^{-1} , I:E ratio 1:2.5–3, peak airway pressure below 25 cmH_2O , variable respiratory rate) to an expired partial pressure of CO_2 between 45 ± 5 mmHg (5.3–6.6 kPa). Artificial ventilation was started immediately after connection to the breathing system and continued until the end of anesthesia. A urinary catheter was placed in all horses immediately following

anesthesia induction and was left in place until the horse had recovered from anesthesia. The amount of urine produced was measured beginning as soon as the urinary catheter was placed until the disconnection of the patient from the large animal anesthesia machine. The urinary production during recovery was not measured. Medetomidine 3.5 $\mu\text{g kg}^{-1} \text{h}^{-1}$ or xylazine 0.69 $\text{mg kg}^{-1} \text{h}^{-1}$ for group MED and group XYL, respectively, were started immediately after positioning the animal on the surgery table, delivered IV by an infusion pump (Phoenix D; Schoch Electronics, Switzerland). A 20 gauge catheter (SURFLO[®], Terumo Europe N.V., Belgium) was placed percutaneously in a transverse facial artery for the measurement of direct arterial blood pressures and to collect blood for arterial blood gas analysis and plasma concentrations of medetomidine and xylazine. A non-distensible extension line filled with heparinized saline solution was connected to the arterial catheter and to an electronic transducer (Monitoring Kit Transpac IT; Abbott Ireland, Ireland) placed at level of shoulder joint (dorsal recumbence) or manubrium sternum (lateral recumbence), zeroed to atmospheric pressure and arterial blood pressures were obtained. HR, respiratory rate (RR), end expiratory carbondioxide tension ($P_{\text{E}}'\text{CO}_2$), expiratory isoflurane concentration ($E_{\text{T}}\text{Iso}$) as well as arterial blood pressures were monitored continuously with a multiparameter monitor (Datex-Ohmeda-CardiCap/5') and manually recorded every 5 min. After anesthesia induction, dobutamine (Dobutrex; Teva Pharma AG, Switzerland) was started at a rate of 40 $\mu\text{g kg}^{-1} \text{h}^{-1}$ via an infusion pump (Volumed[®] μVP5005 ; Arcomed, Switzerland). In order to keep the MAP values stable, the rate of dobutamine was readjusted every 5 min based on a prepared rate scale in dependence on the MAP (<65 mmHg: 70 $\mu\text{g kg}^{-1} \text{h}^{-1}$; 65–70 mmHg: 60 $\mu\text{g kg}^{-1} \text{h}^{-1}$; 71–75 mmHg: 50 $\mu\text{g kg}^{-1} \text{h}^{-1}$; 76–80 mmHg: 40 $\mu\text{g kg}^{-1} \text{h}^{-1}$; 81–85 mmHg: 30 $\mu\text{g kg}^{-1} \text{h}^{-1}$; 86–90 mmHg: 15 $\mu\text{g kg}^{-1} \text{h}^{-1}$; >90 mmHg: 0 $\mu\text{g kg}^{-1} \text{h}^{-1}$). Intravenous crystalloids (Ringer-Laktat; Bichsel AG, Switzerland) were administered IV at a rate of 8–11 $\text{mL kg}^{-1} \text{h}^{-1}$. If despite 70 $\mu\text{g kg}^{-1} \text{h}^{-1}$ dobutamine the MAP remained below 65 mmHg or if the HR increased to a value higher than 1.5 times HR at t_{30} , a bolus of 5 mL kg^{-1} lactated Ringer solution was administered over 10 min. This bolus application was repeated once. If MAP did not rise above 65 mmHg a 2 mL kg^{-1} IV bolus of tetrastarch (Volumen balanced 6%; Fresenius Kabi AG) was administered in addition to the dobutamine infusion. The tetrastarch bolus application was repeated until MAP > 65 mmHg. If nystagmus or incessant fighting against the ventilator occurred, ketamine 0.1 mg kg^{-1} IV was administered. In case of movement, thiopental 0.5 mg kg^{-1} IV (Thiopentalum natricum; Spedalia AG, Switzerland) was given every minute until movement stopped. At time points t_{30} , t_{60} , t_{90} , t_{150} , and t_{210} , arterial blood samples were drawn anaerobically using pre-heparinized syringes (BD A-Line; Becton, Dickinson and Co., UK), and analyzed immediately (RAPIDPoint[®] 500; Siemens, Germany). Analysis included pH, arterial partial pressure of oxygen (PaO_2) and carbon dioxide (PaCO_2), base excess (BE) and electrolytes (Na^+ , K^+ , Ca^{2+} , Cl^-) as well as arterial lactate (Lac). If a PaO_2 < 80 mmHg (10.7 kPa) was detected, FIO_2 was increased to 1.0. If a PaO_2

< 60 mmHg (8 kPa) was detected, salbutamol (Ventolin® GlaxoSmithKline AG, Mûchenbuchsee, Switzerland) $2 \mu\text{g kg}^{-1}$ was administered intratracheally. Thirty minutes prior to the end of anesthesia, morphine 0.1 mg kg^{-1} (Morphine HCL 10 mg; Sintetica SA, Switzerland) was administered IM. At the end of surgery, all infusions and mechanical ventilation were stopped and horses were hoisted from the table to a padded recovery box. From the moment of disconnection of the horse from the anesthetic machine, the recovery timing was started. Ventilation was assisted using a demand valve (JD Medical, Phoenix, Arizona, USA) twice per minute until the horse started to breathe spontaneously. Subsequently, medetomidine $2 \mu\text{g kg}^{-1}$ (group MED) or xylazine 0.3 mg kg^{-1} (group XYL) was administered IV slowly over 2 min. If at that timepoint horses either already moved or showed excessive nystagmus the post anesthesia dose of either medetomidine or xylazine was doubled. Oxygen was administered (15 L min^{-1}) through the endotracheal tube (ETT). The horse's tracheas were extubated 15 min after disconnection of the horse from the anesthetic machine. Before removing the ETT, 5 mL phenylephrine 0.15% drops (Phenylephrin hydrochloridum 1.5 mg mL^{-1} ; Volksapotheke Schaffhausen, Switzerland) were instilled into each nasal cavity. Horses were allowed to recover without assistance. Times to attain sternal and standing positions were noted. The main anesthetist assessed recovery quality. Three different scoring systems (6, 30, 31) were used for assessment.

Plasma medetomidine and xylazine concentrations were determined from blood samples taken from the jugular vein not used for drug injection or (during anesthesia) from the arterial catheter. Blood was sampled before premedication, after sedation, t_{30} , t_{60} , t_{90} , t_{150} , t_{210} , just before the postoperative sedation and 5–10 min after reaching standing position. Before each sampling, 10 mL of blood was withdrawn, then 6 mL of blood was collected into lithium-heparin containing vacutainers (6 mL Vacutainer®, Becton Dickinson, Meylan Cedex, France). This blood was centrifuged without delay and the plasma frozen at -80°C until analyses. Plasma samples were analyzed by liquid chromatography-tandem mass spectrometry (LC-MS/MS). For xylazine determination, a $200 \mu\text{L}$ aliquot of plasma was added with the internal standard (xylazine-d6) and $800 \mu\text{L}$ of acetonitrile, then $100 \mu\text{L}$ of the supernatant were transferred into a vial containing $900 \mu\text{L}$ of 0.1% formic acid aqueous solution prior to injection. As for medetomidine, internal standard medetomidine-d3 was added to $300 \mu\text{L}$ of plasma and a liquid-liquid extraction with $900 \mu\text{L}$ of ethyl acetate was performed. The dry residue of the supernatant was reconstituted in a 0.1% formic acid in water: acetonitrile 90:10 (v/v) solution and injected. The LC system was a Waters Acquity UPLC binary pump, equipped with a Waters BEH C18 ($2.1 \times 50 \text{ mm}$, $1.7 \mu\text{m}$) column (Waters, Milford, MA, USA). The mobile phase was a mixture of acetonitrile and 0.1% formic acid in water, at a flow rate of 0.3 mL min^{-1} under programmed conditions. The detector was a Waters Quattro Premier XE triple quadrupole mass spectrometer (Waters, Milford, MA, USA) with positive electrospray ionization source (ESI+), operating in multiple reaction monitoring (MRM) mode. The specific transitions observed for each analyte were:

221.0 → 89.8 and 163.7 for xylazine; 227.0 → 169.8 for xylazine-d6; 201.0 → 94.7 and 67.7 for medetomidine; 204.0 → 97.6 for medetomidine-d3. The two methods were validated in accordance with EMEA/CHMP/EWP/192217/2009 guidelines before the experiment.

Statistical Analysis

Data distribution was tested with the Shapiro-Wilk test of normality. Depending on distribution, *t*-test or Mann-Whitney test was used to assess differences between groups in terms of weight, age, anesthesia duration, recovery times, intra-anesthetically administered lactated Ringer solution, dobutamine, injected ketamine or thiopental and scores. Repeatedly measured parameters were analyzed using a two-way repeated-measures analysis of variance (ANOVA) followed by Holm-Sidak multiple comparisons test to analyse differences between groups and over time. Chi-square test was used to evaluate significant group differences between required doses of sedation. HR and MAP analyses was restricted to time point $t_{20-t_{120}}$ and $t_{30-t_{120}}$, respectively. Significance was considered when $p \leq 0.05$. Statistical analysis was performed using SigmaStat 3.5 (Systat Software, CA, USA). Results of parametric data are expressed as mean \pm standard error of the mean and results of non-parametric data as median (range).

RESULTS

Sixty horses were randomly allocated into two groups (MED or XYL). Due to a mistake in the arrangement 29 horses were in group XYL and 31 horses in group MED.

One horse in group XYL showed seizures of unknown origin during anesthesia and was completely excluded from data analyses. One horse in group MED needed treatment with salbutamol $2 \mu\text{g kg}^{-1}$ intratracheally but was excluded completely from data analysis due to technical problems 10 min later. Furthermore, one horse from each group had to be excluded from the recovery scoring due to intraoperative decisions for assisted recovery (XYL) or bleeding during recovery (MED). Otherwise, numbers of horses from which data originated are indicated in the corresponding table or figure (Tables 1–3 and Figure 1).

No significant differences in age (MED: 11.5 ± 0.8 ; XYL: 11.1 ± 1.0 years; $p = 0.760$); body weight (MED: 532 ± 14 ; XYL: $520 \pm 19 \text{ kg}$; $p = 0.599$), breed (46 warmbloods; 5 thoroughbreds; 8 ponies), BCS (4 horses BCS 2/5; 47 horses BCS 3/5; 7 horses BCS 4/5; 1 horse BCS 5/5), sex (13 mares; 17 stallions; 29 geldings), type of recumbency (30 dorsal recumbency; 29 lateral recumbency), type of procedure (28 orthopedic procedures; 31 soft tissue procedure) or total duration of anesthesia (MED: 145.4 ± 6.3 ; XYL: $148 \pm 6.8 \text{ min}$; $p = 0.777$) were identified between groups. During preoperative sedation six horses in group XYL showed tumbling before the full dose of xylazine was administered. Further administration of xylazine was stopped immediately and sedation criteria evaluated after 3 min. All six horses pervaded the criteria and anesthesia was induced. In both groups some horses (MED $n = 4$; XYL $n = 2$) did not fulfill the

TABLE 1 | Mean \pm SEM of venous blood-gas values, pH, lactate concentrations, total hemoglobin, base excess in horses with medetomidine isoflurane (group MED) or xylazine isoflurane (group XYL) balanced anesthesia.

Parameter	Group	Time point			
		B	I	R	S
	n MED	31	31	29	29
	n XYL	28	27	28	25
P _v CO ₂ , mmHg (kPa)	MED	46.89 \pm 0.73 (6.3 \pm 0.1)	48.20 \pm 0.73 (6.4 \pm 0.1)	62.27 \pm 0.76 (8.3 \pm 0.1) [#]	52.11 \pm 0.76 (6.9 \pm 0.1) [#]
	XYL	46.58 \pm 0.77 (6.2 \pm 0.1)	49.14 \pm 0.78 (6.6 \pm 0.1) [#]	63.02 \pm 0.77 (8.4 \pm 0.1) [#]	53.54 \pm 0.82 (7.1 \pm 0.1) [#]
P _v O ₂ , mmHg (kPa)	MED	34.35 \pm 0.80 (4.6 \pm 0.1)	28.44 \pm 0.81 (3.8 \pm 0.1) [#]	47.62 \pm 0.83 (6.3 \pm 0.1) ^{*#}	33.64 \pm 0.83 (4.5 \pm 0.1)
	XYL	34.90 \pm 0.83 (4.7 \pm 0.1)	28.33 \pm 0.85 (3.8 \pm 0.1) [#]	44.90 \pm 0.83 (6.0 \pm 0.1) ^{*#}	33.85 \pm 0.90 (4.5 \pm 0.1)
pH	MED	7.41 \pm 0.004	7.39 \pm 0.004 [#]	7.32 \pm 0.004 [#]	7.40 \pm 0.004 [#]
	XYL	7.41 \pm 0.004	7.38 \pm 0.004 [#]	7.32 \pm 0.004 [#]	7.39 \pm 0.004 [#]
Lactate (mmol L ⁻¹)	MED	0.63 \pm 0.16	0.69 \pm 0.16	2.02 \pm 0.17 [#]	2.03 \pm 0.17 [#]
	XYL	0.62 \pm 0.17	0.70 \pm 0.17	1.58 \pm 0.17 [#]	1.80 \pm 0.19 [#]
Hb (g dL ⁻¹)	MED	12.61 \pm 0.21	12.11 \pm 0.20	10.99 \pm 0.22 [#]	11.60 \pm 0.22 [#]
	XYL	12.61 \pm 0.22	11.69 \pm 0.22 [#]	10.68 \pm 0.22 [#]	12.06 \pm 0.23
BE (mmol L ⁻¹)	MED	4.00 \pm 0.29	2.64 \pm 0.29 [#]	4.01 \pm 0.30	5.72 \pm 0.30 [#]
	XYL	3.41 \pm 0.30	2.02 \pm 0.31 [#]	4.43 \pm 0.30 [#]	5.90 \pm 0.32 [#]

Significant differences ($p < 0.05$) between groups are indicated by * and significant changes compared to B in the same group are indicated by #.

P_vO₂, P_vCO₂, venous oxygen and carbon dioxide partial pressures; Lac, lactate concentration; Hb, total hemoglobin; BE, base excess; B, Baseline; I, Induction; R, Recovery; S, Standing.

TABLE 2 | Mean \pm SEM of arterial blood-gas values, pH, lactate concentrations, total hemoglobin, base excess in horses with medetomidine isoflurane (group MED) or xylazine isoflurane (group XYL) balanced anesthesia.

Parameter	Group	Time point			
		t ₃₀	t ₆₀	t ₉₀	t ₁₅₀
	n MED	31	30	26	8
	n XYL	27	27	26	10
P _a CO ₂ , mmHg (kPa)	MED	50.57 \pm 0.49 (6.7 \pm 0.07)	52.89 \pm 0.51 (7.1 \pm 0.07) ^{*#}	54.82 \pm 0.56 (7.3 \pm 0.07) ^{*#}	55.56 \pm 1.10 (7.4 \pm 0.15) [#]
	XYL	51.19 \pm 0.52 (6.8 \pm 0.07)	51.03 \pm 0.54 (6.8 \pm 0.07) [*]	52.11 \pm 0.55 (6.9 \pm 0.07) [*]	52.45 \pm 0.96 (7.0 \pm 0.13)
P _a O ₂ , mmHg (kPa)	MED	153.97 \pm 5.16 (20.5 \pm 0.69)	151.34 \pm 5.45 (20.2 \pm 0.73)	146.47 \pm 5.90 (19.5 \pm 0.79)	152.00 \pm 11.40 (20.3 \pm 1.52)
	XYL	151.03 \pm 5.53 (20.1 \pm 0.74)	148.56 \pm 5.53 (19.8 \pm 0.79)	155.82 \pm 5.68 (20.8 \pm 0.76)	145.30 \pm 10.00 (19.4 \pm 1.33)
pH	MED	7.35 \pm 0.003	7.35 \pm 0.003	7.35 \pm 0.003	7.36 \pm 0.006
	XYL	7.35 \pm 0.003	7.36 \pm 0.003	7.36 \pm 0.003	7.38 \pm 0.006 [#]
Lactate (mmol L ⁻¹)	MED	1.61 \pm 0.03	1.83 \pm 0.03 [#]	1.9 \pm 0.03 [#]	2.00 \pm 0.06 [#]
	XYL	1.47 \pm 0.03	1.60 \pm 0.03 [#]	1.65 \pm 0.03 [#]	1.56 \pm 0.06 [#]
Hb (g dL ⁻¹)	MED	11.32 \pm 0.12	10.6 \pm 0.12 [#]	10.40 \pm 0.13 [#]	10.88 \pm 0.25
	XYL	11.62 \pm 0.12	10.76 \pm 0.13 [#]	10.35 \pm 0.13 [#]	9.52 \pm 0.24 [#]
BE (mmol L ⁻¹)	MED	2.37 \pm 0.20	2.00 \pm 0.21 [#]	2.87 \pm 0.23 [#]	4.55 \pm 0.44 [#]
	XYL	1.56 \pm 0.21	2.10 \pm 0.22	2.50 \pm 0.22 [#]	4.05 \pm 0.39 [#]

Significant differences ($p < 0.05$) between groups are indicated by * and significant changes compared to t₃₀ in the same group are indicated by #.

P_aO₂, P_aCO₂, arterial oxygen and carbon dioxide partial pressures; Lac, lactate concentration; Hb, total hemoglobin; BE, base excess; t_{30–150}, timepoints in minutes after induction.

sedation criteria and required one or more supplemental dose. The median dose of sedation necessary to fulfill sedation criteria prior to anesthesia induction was 7 (7–10) $\mu\text{g kg}^{-1}$ in group MED and 1.1 (0.8–1.3) mg kg^{-1} in group XYL. There was no significant difference in the time from the start of sedation to induction of anesthesia between groups [MED: 6 (3–8); XYL: 7 (4–14) min; $p = 0.101$].

No significant difference was found between groups regarding mean E_T'Iso concentrations (MED: 1.1 \pm 0.02; XYL: 1.1 \pm

0.02%; $p = 0.267$). Nevertheless, group-independent changes over time were significant with E_T'ISO increasing over time compared to t₂₀.

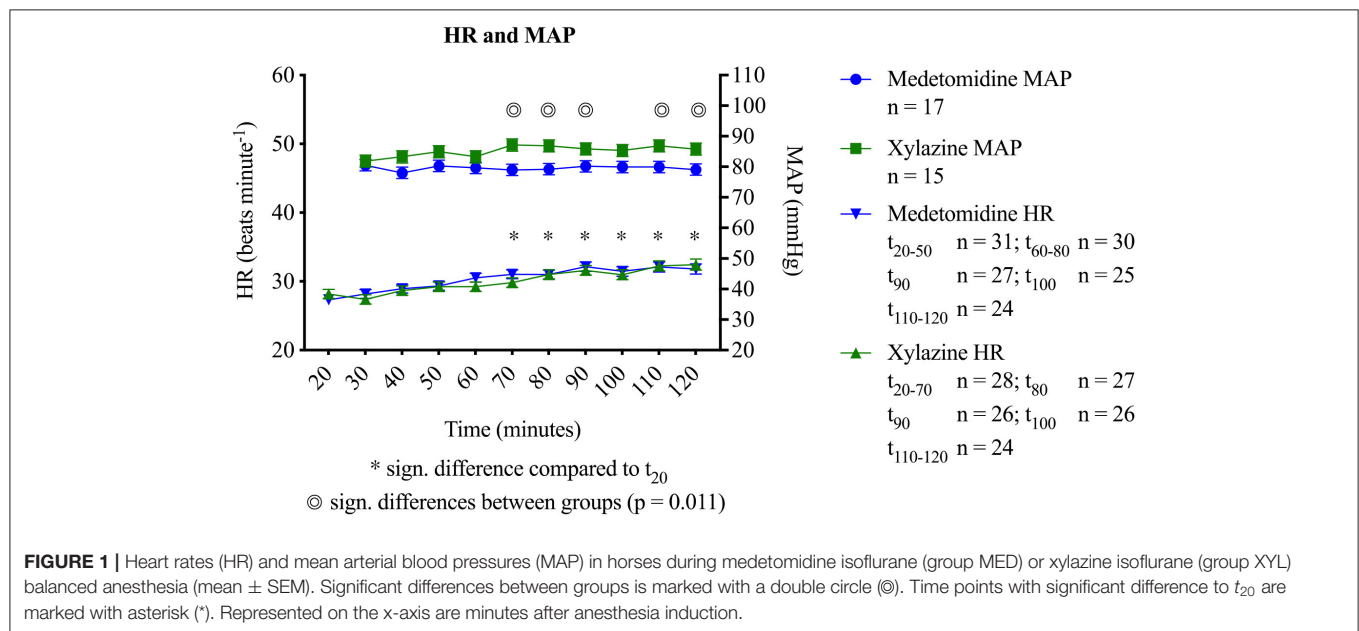
Figure 1 displays MAP and HR. MAP was within the targeted values but significantly higher with xylazine compared to medetomidine at time points t₇₀ ($p = 0.007$), t₈₀ ($p = 0.005$), t₉₀ ($p = 0.017$), t₁₁₀ ($p = 0.013$) and t₁₂₀ ($p = 0.015$). No differences between groups were detected for HR ($p = 0.85$). However, an increase in HR over time was observed for both

TABLE 3 | Mean \pm SEM of electrolytes in horses with medetomidine isoflurane (group MED) or xylazine isoflurane (group XYL) balanced anesthesia.

Parameter	Group	Time point							
		B (venous)	I (venous)	t ₃₀ (arterial)	t ₆₀ (arterial)	t ₉₀ (arterial)	t ₁₅₀ (arterial)	R (venous)	S (venous)
	n MED	31	31	31	30	26	8	29	29
	n XYL	28	27	27	27	26	10	28	25
Na ⁺ (mmol L ⁻¹)	MED	135.99 \pm 0.31	135.94 \pm 0.30					135.09 \pm 0.31	135.83 \pm 0.31
	XYL	135.81 \pm 0.32	136.89 \pm 0.32					134.99 \pm 0.31 [#]	136.77 \pm 0.34
K ⁺ (mmol L ⁻¹)	MED	3.58 \pm 0.04	3.49 \pm 0.04					3.71 \pm 0.04 [#]	3.91 \pm 0.04 [#]
	XYL	3.67 \pm 0.04	3.55 \pm 0.04					3.68 \pm 0.04	3.98 \pm 0.04 [#]
Ca ²⁺ (mmol L ⁻¹)	MED*	1.46 \pm 0.007	1.45 \pm 0.007					1.30 \pm 0.007 ^{*#}	1.28 \pm 0.007 ^{*#}
	XYL*	1.49 \pm 0.007	1.48 \pm 0.007					1.34 \pm 0.007 ^{*#}	1.31 \pm 0.007 ^{*#}
Cl ⁻ (mmol L ⁻¹)	MED	100.45 \pm 0.24	100.58 \pm 0.24					99.31 \pm 0.25 [#]	98.24 \pm 0.25 [#]
	XYL	100.39 \pm 0.25	100.44 \pm 0.26					99.18 \pm 0.25 [#]	98.28 \pm 0.27 [#]
Na ⁺ (mmol L ⁻¹)	MED			134 \pm 0.25	134.04 \pm 0.26	134.51 \pm 0.30 [#]	134.54 \pm 0.56 [#]		
	XYL			133.26 \pm 0.27	133.64 \pm 0.28	134.32 \pm 0.29	134.15 \pm 0.50		
K ⁺ (mmol L ⁻¹)	MED			3.6 \pm 0.02	3.5 \pm 0.02	3.50 \pm 0.03	3.69 \pm 0.05		
	XYL			3.54 \pm 0.02	3.53 \pm 0.03	3.53 \pm 0.03	3.52 \pm 0.05		
Ca ²⁺ (mmol L ⁻¹)	MED			1.38 \pm 0.005	1.34 \pm 0.005 [#]	1.32 \pm 0.006 [#]	1.29 \pm 0.011 [#]		
	XYL			1.4 \pm 0.005	1.36 \pm 0.006 [#]	1.34 \pm 0.006 [#]	1.32 \pm 0.010 [#]		
Cl ⁻ (mmol L ⁻¹)	MED			99.77 \pm 0.20	100.10 \pm 0.20	100.12 \pm 0.22	100.13 \pm 0.43		
	XYL			99.61 \pm 0.20	100.22 \pm 0.21 [#]	100.19 \pm 0.22	101.00 \pm 0.38 [#]		

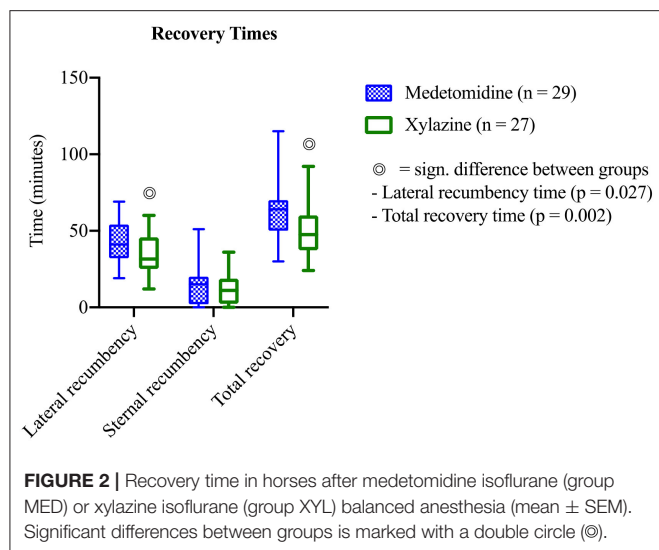
Significant differences ($p < 0.05$) between groups are indicated by * and significant changes compared to B or t₃₀ in the same group are indicated by #.

Na⁺, ionized sodium; K⁺, ionized potassium; Ca²⁺, ionized calcium; Cl⁻, ionized chlorine; B, Baseline; I, Induction; R, Recovery; S, Standing; t_{30–150}, timepoints in minutes after induction.



groups ($p < 0.001$). Dobutamine requirement was significantly higher with medetomidine (MED: 36 ± 1.8 ; XYL: 24 ± 1.8 $\mu\text{g kg}^{-1} \text{ h}^{-1}$; $p = 0.0003$). Five horses in group MED required between one and three lactated Ringer solution boluses and one of them needed two boluses of tetrastarch, while in group XYL only three horses received one bolus of lactated Ringer solution to achieve the targeted values of MAP and

HR. Results of venous and arterial pH and arterial blood gases are found in **Tables 1, 2** and those of electrolytes, lactate and hemoglobin in **Table 3**. There were some changes over time and very minimal differences in arterial partial pressure of CO₂ (P_aCO₂), venous partial pressure of O₂ (P_vO₂), lactate, hemoglobin concentrations and Ca²⁺ between the groups (**Tables 1–3**).



Multiple horses in both groups (MED $n = 18$; XYL $n = 15$) required one or several boluses of ketamine. Two horses in MED and three horses in XYL were administered a single dose of 0.5 or 1.0 mg kg^{-1} thiopental, either to allow intubation (MED $n = 2$; XYL $n = 2$) or because of limb movement during surgery (XYL $n = 1$). There was no significant difference in urine production between the two groups [MED: $6.4 (1.6\text{--}14.6)$; XYL: $5.0 (1.6\text{--}15.4) \text{ mL kg}^{-1} \text{ h}^{-1}$; $p = 0.522$].

All horses started to breathe spontaneously within 3–4 min after disconnection from the breathing system and post sedation was administered right thereafter. Total recovery time was significantly longer with medetomidine [MED: $64 (30\text{--}115)$; XYL: $47.5 (24\text{--}92) \text{ min}$; $p = 0.002$] (Figure 2). The pivotal phase during recovery was lateral recumbency time, which was significantly longer with medetomidine [MED: $41 (19\text{--}69)$; XYL: $31.5 (12\text{--}60) \text{ min}$; $p = 0.027$]. No significant difference between groups in the other recorded time spans were detected (in minutes): time to extubation [MED: $15 (11\text{--}40)$; XYL: $15 (8\text{--}30)$; $p = 0.114$]; time from sternal to standing [MED: $15 (0\text{--}51)$; XYL: $11 (0\text{--}36)$; $p = 0.615$]. Recovery was generally of good quality and showed no significant differences between groups in all three scoring systems. In score Sacks et al. (6) where one is best and five is worst recovery quality, 16 horses of group MED and 14 horses of group XYL were classified score 1. The worst recovery scores recorded were from three horses in group MED with a score of 3 and one horse in group XYL with a score of 4 [MED: $1.6 (1\text{--}3)$; XYL: $1.7 (1\text{--}4)$; $p = 0.837$]. In the scoring system by Valverde et al. (31) where recovery quality can range between 8 and 26 plus number of attempts to stand scores were MED: $14 (9\text{--}25)$; XYL: $12 (9\text{--}27)$; $p = 0.627$. Scores of the third scoring system (30) used can range from 8 to 70 plus number of attempts to sternal plus number of attempts to stand [MED: $23 (12\text{--}56)$; XYL: $24 (10\text{--}52)$; $p = 0.712$]. Two horses, one of each group, were administered an additional bolus of either medetomidine or xylazine for recovery, as they showed rapid nystagmus upon arrival in the recovery box

which did not cease following the administration of the normal sedative dose.

Plasma concentrations of both drugs showed a constant level during general anesthesia (Figure 3).

DISCUSSION

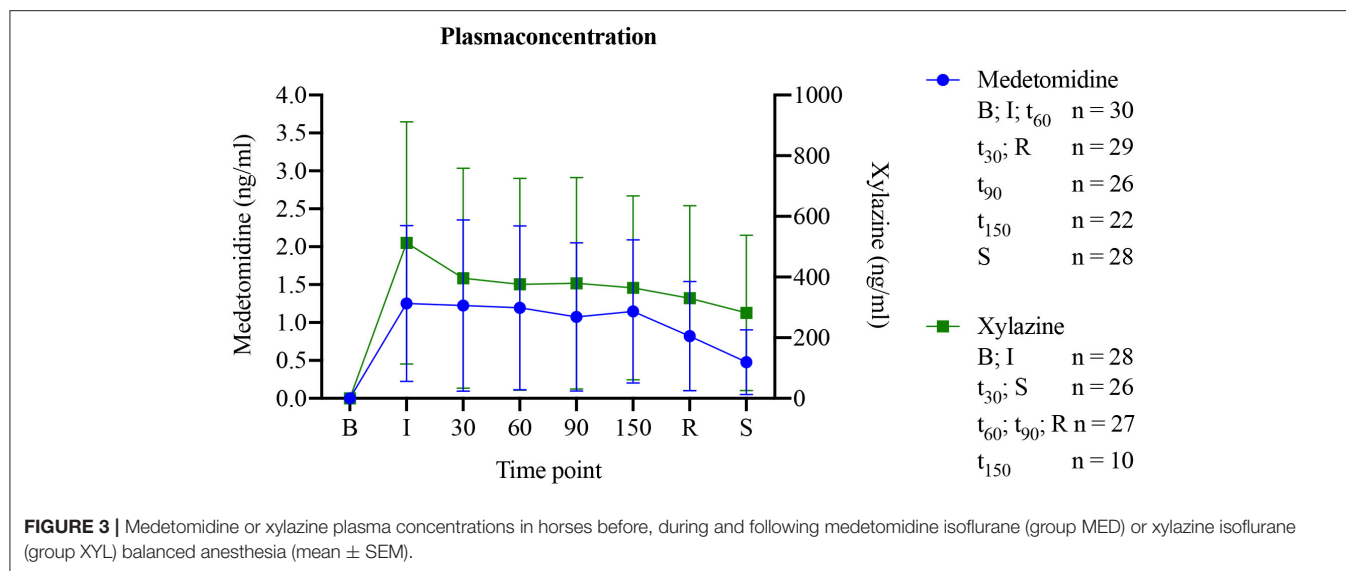
The current study investigated in horses anesthetized with isoflurane whether the use of medetomidine compared to xylazine for balanced anesthesia resulted in clinically relevant differences in cardiopulmonary function or recovery from anesthesia. The only detectable clinically relevant difference was a shorter recovery following the use of xylazine. Further at some timepoints slightly lower mean arterial blood pressures in combination with a lower need for dobutamine with xylazine were observed. As arterial blood pressures were easily maintained within clinically normal values, the relevance of this finding yet has to be determined.

There is no doubt that even with modern anesthesia techniques and optimized monitoring systems fatality rate in horses is too high and studies to establish safer anesthesia techniques are warranted. Since most anesthetists have stopped to use halothane and use isoflurane or sevoflurane instead (32), less horses have been reported to die intraoperatively, but fatal problems during recovery are predominant (2, 33). Thus, anesthesia techniques resulting in calm recovery should be established without delay.

Alpha₂-adrenoceptor agonists based PIVA techniques have been used since about two decades (34, 35) and in particular medetomidine and more recently dexmedetomidine have been tested in many horse studies (6, 10, 12). As those drugs are short acting (19, 36, 37) they can be easily titrated to effect and recovery, even after prolonged anesthesia, is usually prompt and of better quality than following lidocaine or ketamine based PIVA protocols (12, 38). As in European countries neither medetomidine nor dexmedetomidine are licensed for food producing animals PIVA protocols including licensed alpha₂-adrenoceptor agonists are warranted. In conscious horses xylazine, in comparison to the other two for horses licensed alpha₂-adrenoceptor agonists detomidine and romifidine, has a similar short duration of action as medetomidine and dexmedetomidine (20, 39, 40). Also, distribution half-lives of xylazine are comparable to medetomidine and dexmedetomidine (15, 41). Romifidine and detomidine are longer acting and show more cardiovascular effects than xylazine (20, 42).

We chose to compare medetomidine with xylazine as dexmedetomidine for sedation of clinical patients proven to be not as reliable as medetomidine (6, 43) and dose rates necessary to achieve good sedation before anesthesia induction were considered too variable to be a safe alternative for daily practice.

The present study detected some differences in cardiopulmonary function between the two regimes tested. If the higher arterial blood pressures with xylazine are a result of more vasoconstriction or a better cardiac output will have to be tested in a study with detailed monitoring of cardiopulmonary function. In conscious horses with xylazine at the same dose as



used in the present study, effects on cardiopulmonary function were remarkably low (42). It is however noteworthy that with a very low dose dobutamine MAP stayed within targeted values in both groups, but in the XYL group about 30% less dobutamine was needed. We did not use LIDCO to measure cardiac output even though it would be the only safe method to measure cardiac output in clinical cases, as several of the drugs used in the present study have experimentally been shown to have an influence on the measurements of LIDCO (44).

In conscious horses and ponies xylazine and medetomidine at the dose rates used in the current study, cause significant cardiopulmonary depression following bolus administration (19, 20, 42). However, at steady state plasma levels cardiopulmonary function returns to nearly pre-sedation values (19, 42). The plasma levels of both drugs were stable during drug infusion and remarkably similar to the plasma levels measured in horses sedated with similar dose rates of xylazine (26) and slightly higher than what has been reported in standing ponies (19). Although in both groups at the end of CRI following transport into recovery a relevant bolus dose was given to keep the horses sedated and prevent them from an untimely recovery, plasma levels showed a rapid decline, probably as a result of a fast decrease of isoflurane coupled to better liver and general perfusion influencing volume of drug distribution and metabolism.

Acid base balance, electrolytes, lactate and glucose concentrations as well as arterial blood gases remained within clinically acceptable ranges and differences don't seem to be clinically relevant. The amount of urine produced in both groups was considerable and needs to be taken into consideration, when fluid therapy is planned. The use of an intraurethral catheter is advisable in any case.

Bryant et al. (39) showed that medetomidine produces more ataxia than xylazine, so a more ataxic recovery might be expected following medetomidine CRI. Bryant et al. (39) tested medetomidine at a considerably higher dose rate (MED 10 $\mu\text{g kg}^{-1}$ IV) than what was used here for recovery. Therefore,

recovery quality in the present study was not different between the groups, as sedative doses used were adequate and not causing additional ataxia. We used 3 different previously described scoring systems (6, 30, 31) to judge recovery as each system evaluates recovery in a slightly different way. We wanted to ascertain that our results are not a consequence of the scoring system used but rather of the medication tested. The groups were similar concerning factors that might affect recovery quality such as anesthesia duration, weight and recumbency during surgery (45, 46). As the incidence of poor recoveries in this study was low and no ASA III-IV horses were included we cannot conclude with only 56 observed recoveries, if the use of either drug is really equal concerning recovery quality. A power analyses of our data revealed that a total of 498 comparable horse anesthetics would have to be investigated to detect with a power of 80% and an alpha error of 0.05, if the incidence of two bad quality recoveries [score 4 and 5 in comparison to 1, 2, or 3 according to Sacks et al. (6)] can be reduced to one, with one or the other PIVA regime.

With xylazine in our study horses remained for a shorter time period in lateral recumbency and recovered overall faster. We can only speculate if this is a result of a quicker redistribution or metabolism of xylazine or a stronger sedative/analgesic effect of the more specific α_2 -adrenoceptor agonists medetomidine. Theoretically, recoveries are of better quality if horses don't get up too quickly, as it allows for the inhalant anesthetic to be blown off before the horse attempts to stand up. But with the shorter acting dexmedetomidine recovery from balanced anesthesia was of better quality than following medetomidine PIVA (6), despite otherwise more or less identical effects of the two drugs (at equipotent doses). On the other hand, will prolonged recumbency cause atelectasis and poor perfusion of dependent muscle groups resulting in myopathy and neuropathy. Therefore, the use of xylazine, in particular for prolonged anesthesia's, in compromised patients might prove to be advantageous.

The major limitation of this study is that no sample size calculation with a main outcome was performed. A further weakness is that the depth of anesthesia was judged by a subjective method, but the anesthetist was the same for all procedures and was blinded to treatment. Also, for different procedures horses received two different non-steroidal anti-inflammatories, flunixin and phenylbutazone, but this was equally distributed to the groups.

CONCLUSION

In conclusion PIVA with isoflurane and a CRI of xylazine ($0.69 \text{ mg kg}^{-1} \text{ h}^{-1}$) compared to medetomidine ($3.5 \text{ } \mu\text{g kg}^{-1} \text{ h}^{-1}$) resulted in slightly better MAP, lower need for dobutamine and a faster recovery, whilst overall recovery quality was unaffected.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

REFERENCES

1. Steffey EP, Howland D Jr. Cardiovascular effects of halothane in the horse. *Am J Vet Res.* (1978) 39:611–5.
2. Dugdale AH, Taylor PM. Equine anaesthesia-associated mortality: where are we now? *Vet Anaesth Analg.* (2016) 43:242–55. doi: 10.1111/vaa.12372
3. Johnston GM, Eastment JK, Wood J, Taylor PM. The confidential enquiry into perioperative equine fatalities (CEPEF): mortality results of Phases 1 and 2. *Vet Anaesth Analg.* (2002) 29:159–70. doi: 10.1046/j.1467-2995.2002.00106.x
4. Gozalo-Marcilla M, Gasthuys F, Schauvliege S. Partial intravenous anaesthesia in the horse: a review of intravenous agents used to supplement equine inhalation anaesthesia. Part 1: lidocaine and ketamine. *Vet Anaesth Analg.* (2014) 41:335–45. doi: 10.1111/vaa.12179
5. Gozalo-Marcilla M, Gasthuys F, Schauvliege S. Partial intravenous anaesthesia in the horse: a review of intravenous agents used to supplement equine inhalation anaesthesia. Part 2: opioids and alpha-2 adrenoceptor agonists. *Vet Anaesth Analg.* (2015) 42:1–16. doi: 10.1111/vaa.12196
6. Sacks M, Ringer SK, Bischofberger AS, Berchtold SM, Bettschart-Wolfensberger R. Clinical comparison of dexmedetomidine and medetomidine for isoflurane balanced anaesthesia in horses. *Vet Anaesth Analg.* (2017) 44:1128–38. doi: 10.1016/j.vaa.2016.12.061
7. Nannarone S, Spadavecchia C. Evaluation of the clinical efficacy of two partial intravenous anesthetic protocols, compared with isoflurane alone, to maintain general anesthesia in horses. *Am J Vet Res.* (2012) 73:959–67. doi: 10.2460/ajvr.73.7.959
8. Bettschart-Wolfensberger R, Larenza Menzies MP. Balanced anesthesia in the equine. *Clin Tech Equine Pract.* (2007) 6:104–10. doi: 10.1053/j.ctep.2007.05.002
9. Devisscher L, Schauvliege S, Dewulf J, Gasthuys F. Romifidine as a constant rate infusion in isoflurane anesthetized horses: a clinical study. *Vet Anaesth Analg.* (2010) 37:425–33. doi: 10.1111/j.1467-2995.2010.00556.x
10. Marcilla MG, Schauvliege S, Segaeert S, Duchateau L, Gasthuys F. Influence of a constant rate infusion of dexmedetomidine on cardiopulmonary function and recovery quality in isoflurane anesthetized horses. *Vet Anaesth Analg.* (2012) 39:49–58. doi: 10.1111/j.1467-2995.2011.00672.x

ETHICS STATEMENT

The animal study was reviewed and approved by ZH Veterinäramt Zürich. Written informed consent was obtained from the owners for the participation of their animals in this study.

AUTHOR CONTRIBUTIONS

AW: study design, acquisition, analysis and interpretation of data and the writing of the manuscript. RB-W: supervised the study and contributed to every part of it. AB: analysis of plasma samples. SR: statistical analysis and interpretation of data. AB, FJ, and MB: collection of data and supervision of clinical study progress. FJ: responsible for randomization of the patients. All authors contributed to the critical revision of the manuscript and approved the final manuscript for publication.

ACKNOWLEDGMENTS

The authors thank the assistants of the Equine Department, who helped with the organization of this study, namely, with patient recruitment and obtaining owner's consents.

11. Poppel N, Hopster K, Geburek F, Kastner S. Influence of ketamine or xylazine supplementation on isoflurane anesthetized horses—a controlled clinical trial. *Vet Anaesth Analg.* (2015) 42:30–8. doi: 10.1111/vaa.12176
12. Ringer SK, Kalchhofner K, Boller J, Furst A, Bettschart-Wolfensberger R. A clinical comparison of two anaesthetic protocols using lidocaine or medetomidine in horses. *Vet Anaesth Analg.* (2007) 34:257–68. doi: 10.1111/j.1467-2995.2006.00321.x
13. Schauvliege S, Marcilla MG, Verryken K, Duchateau L, Devisscher L, Gasthuys F. Effects of a constant rate infusion of detomidine on cardiovascular function, isoflurane requirements and recovery quality in horses. *Vet Anaesth Analg.* (2011) 38:544–54. doi: 10.1111/j.1467-2995.2011.00659.x
14. Grimsrud KN, Mama KR, Steffey EP, Stanley SD. Pharmacokinetics and pharmacodynamics of intravenous medetomidine in the horse. *Vet Anaesth Analg.* (2012) 39:38–48. doi: 10.1111/j.1467-2995.2011.00669.x
15. Garcia-Villar R, Toutain PL, Alvinerie M, Ruckebusch Y. The pharmacokinetics of xylazine hydrochloride: an interspecific study. *J Vet Pharmacol Ther.* (1981) 4:87–92. doi: 10.1111/j.1365-2885.1981.tb00715.x
16. Virtanen R, Savola JM, Saano V, Nyman L. Characterization of the selectivity, specificity and potency of medetomidine as an alpha 2-adrenoceptor agonist. *Eur J Pharmacol.* (1988) 150:9–14. doi: 10.1016/0014-2999(88)90744-3
17. Kamerling S, Keowen M, Bagwell C. Pharmacological profile of medetomidine in the equine. *Acta Vet Scand.* (1991) 161–2.
18. Creighton CM, Lemke KA, Lamont LA, Horney BS, Riley CB. Comparison of the effects of xylazine bolus versus medetomidine constant rate infusion on cardiopulmonary function and depth of anesthesia in horses anesthetized with isoflurane. *J Am Vet Med Assoc.* (2012) 240:991–7. doi: 10.2460/javma.240.8.991
19. Bettschart-Wolfensberger R, Bettschart RW, Vainio O, Marlin D, Clarke KW. Cardiopulmonary effects of a two hour medetomidine infusion and its antagonism by atipamezole in horses and ponies. *J Vet Anaesth.* (1999) 26:8–12. doi: 10.1111/j.1467-2995.1999.tb00176.x
20. Yamashita K, Tsubakishita S, Futakoshi S, Ueda I, Hamaguchi H, Seno T, et al. Cardiovascular effects of medetomidine, detomidine and xylazine in horses. *J Vet Med Sci.* (2000) 62:1025–32. doi: 10.1292/jvms.62.1025
21. Kalchhofner KS, Picek S, Ringer SK, Jackson M, Hassig M, Bettschart-Wolfensberger R. A study of cardiovascular function under controlled and

- spontaneous ventilation in isoflurane-medetomidine anaesthetized horses. *Vet Anaesth Analg.* (2009) 36:426–35. doi: 10.1111/j.1467-2995.2009.00477.x
22. Yamashita K, Muir WW 3rd, Tsubakishita S, Abrahamsen E, Lerch P, Hubbell JA, et al. Clinical comparison of xylazine and medetomidine for premedication of horses. *J Am Vet Med Assoc.* (2002) 221:1144–9. doi: 10.2460/javma.2002.221.1144
 23. Menzies MP, Ringer SK, Conrot A, Theurillat R, Kluge K, Kutter AP, et al. Cardiopulmonary effects and anaesthesia recovery quality in horses anaesthetized with isoflurane and low-dose S-ketamine or medetomidine infusions. *Vet Anaesth Analg.* (2016) 43:623–34. doi: 10.1111/vaa.12359
 24. Daunt DA, Steffey EP. Alpha-2 adrenergic agonists as analgesics in horses. *Vet Clin North Am Equine Pract.* (2002) 18:39–46, vi. doi: 10.1016/S0749-0739(02)00004-4
 25. England GC, Clarke KW. Alpha 2 adrenoceptor agonists in the horse—a review. *Br Vet J.* (1996) 152:641–57. doi: 10.1016/S0007-1935(96)80118-7
 26. Ringer SK, Portier KG, Fourel I, Bettschart-Wolfensberger R. Development of a xylazine constant rate infusion with or without butorphanol for standing sedation of horses. *Vet Anaesth Analg.* (2012) 39:1–11. doi: 10.1111/j.1467-2995.2011.00653.x
 27. Bryant CE, Clarke KW, Thompson J. Cardiopulmonary effects of medetomidine in sheep and in ponies. *Res Vet Sci.* (1996) 60:267–71. doi: 10.1016/S0034-5288(96)90052-1
 28. Steffey EP, Pascoe PJ, Woliner MJ, Berryman ER. Effects of xylazine hydrochloride during isoflurane-induced anesthesia in horses. *Am J Vet Res.* (2000) 61:1225–31. doi: 10.2460/ajvr.2000.61.1225
 29. Taylor P, Coumbe K, Henson F, Scott D, Taylor A. Evaluation of sedation for standing clinical procedures in horses using detomidine combined with buprenorphine. *Vet Anaesth Analg.* (2014) 41:14–24. doi: 10.1111/vaa.12055
 30. Leece EA, Corletto F, Brearley JC. A comparison of recovery times and characteristics with sevoflurane and isoflurane anaesthesia in horses undergoing magnetic resonance imaging. *Vet Anaesth Analg.* (2008) 35:383–91. doi: 10.1111/j.1467-2995.2008.00399.x
 31. Valverde A, Black B, Cribb NC, Hathway A, Daw A. Assessment of unassisted recovery from repeated general isoflurane anesthesia in horses following post-anesthetic administration of xylazine or acepromazine or a combination of xylazine and ketamine. *Vet Anaesth Analg.* (2013) 40:3–12. doi: 10.1111/j.1467-2995.2012.00782.x
 32. Wohlfender FD, Doherr MG, Driessen B, Hartnack S, Johnston GM, Bettschart-Wolfensberger R. International online survey to assess current practice in equine anaesthesia. *Equine Vet J.* (2015) 47:65–71. doi: 10.1111/evj.12257
 33. Laurenza C, Ansart L, Portier K. Risk factors of anesthesia-related mortality and morbidity in one equine hospital: a retrospective study on 1,161 cases undergoing elective or emergency surgeries. *Front Vet Sci.* (2019) 6:514. doi: 10.3389/fvets.2019.00514
 34. Bettschart-Wolfensberger R, Jaggin-Schmucker N, Lendl C, Bettschart RW, Clarke KW. Minimal alveolar concentration of desflurane in combination with an infusion of medetomidine for the anaesthesia of ponies. *Vet Rec.* (2001) 148:264–7. doi: 10.1136/vr.148.9.264
 35. Kalchofner KS, Ringer SK, Boller J, Kästner SBR, Lischer C, Bettschart-Wolfensberger R. Clinical assessment of anesthesia with isoflurane and medetomidine in 300 equidae. *Pferdeheilkunde.* (2006) 22:301–8. doi: 10.21836/PEM20060309
 36. Bettschart-Wolfensberger R, Clarke KW, Vainio O, Aliabadi F, Demuth D. Pharmacokinetics of medetomidine in ponies and elaboration of a medetomidine infusion regime which provides a constant level of sedation. *Res Vet Sci.* (1999) 67:41–6. doi: 10.1053/rvsc.1998.0274
 37. Bettschart-Wolfensberger R, Freeman SL, Bowen IM, Aliabadi FS, Weller R, Huhtinen M, et al. Cardiopulmonary effects and pharmacokinetics of i.v. dexmedetomidine in ponies. *Equine Vet J.* (2005) 37:60–4. doi: 10.2746/0425164054406801
 38. Larenza MP, Ringer SK, Kutter AP, Conrot A, Theurillat R, Kummer M, et al. Evaluation of anesthesia recovery quality after low-dose racemic or S-ketamine infusions during anesthesia with isoflurane in horses. *Am J Vet Res.* (2009) 70:710–8. doi: 10.2460/ajvr.70.6.710
 39. Bryant CE, England GCW, Clarke KW. A comparison of the sedative effects of medetomidine and xylazine in the horse. *Vet Anaesth Analg.* (1991) 18:55–7. doi: 10.1111/j.1467-2995.1991.tb00511.x
 40. Bueno AC, Cornick-Seahorn J, Seahorn TL, Hosgood G, Moore RM. Cardiopulmonary and sedative effects of intravenous administration of low doses of medetomidine and xylazine to adult horses. *Am J Vet Res.* (1999) 60:1371–6.
 41. Nych HK, Stanley SD, McKemie DS, Arthur RM, Kass PH. Pharmacokinetic and pharmacodynamics of xylazine administered to exercised thoroughbred horses. *Drug Test Anal.* (2017) 9:713–20. doi: 10.1002/dta.2047
 42. Ringer SK, Schwarzwald CC, Portier KG, Ritter A, Bettschart-Wolfensberger R. Effects on cardiopulmonary function and oxygen delivery of doses of romifidine and xylazine followed by constant rate infusions in standing horses. *Vet J.* (2013) 195:228–34. doi: 10.1016/j.tvjl.2012.06.036
 43. Gozalo-Marcilla M, Gasthuys F, Luna SPL, Schauvliege S. Is there a place for dexmedetomidine in equine anaesthesia and analgesia? A systematic review. (2005–2017). *J Vet Pharmacol Ther.* (2018) 41:205–17. doi: 10.1111/jvp.12474
 44. Ambrisko TD, Kabes R, Moens Y. Influence of drugs on the response characteristics of the LiDCO sensor: an in vitro study. *Br J Anaesth.* (2013) 110:305–10. doi: 10.1093/bja/aes380
 45. Hubbell J, Muir W. Considerations for induction, maintenance, and recovery. *Equine Anesth.* (2009) 381–96. doi: 10.1016/B978-1-4160-2326-5.00021-3
 46. Wagner AE. Complications in equine anesthesia. *Vet Clin North Am Equine Pract.* (2008) 24:735–52, x. doi: 10.1016/j.cveq.2008.10.002

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Wiederkehr, Barbarossa, Ringer, Jörgen, Bryner and Bettschart-Wolfensberger. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.